

SHORT COMMUNICATION

Identification of CYP2B14P and CYP2B16P, Two Apparent Pseudogenes in the Rat Cytochrome P450 2B (CYP2B) Subfamily

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ABSTRACT. Cytochrome P450 2B3 (CYP2B3) is a member of the CYP2B subfamily and is present constitutively in rat liver. During cloning of the CYP2B3 gene, several variant genomic inserts were isolated, exoncontaining fragments of which were identified by hybridizing to CYP2B3 cDNA or exonic probes. Nucleotide sequence analysis of one set of variant inserts showed them to carry exon 1 and the 5'-flanking region of a gene designated CYP2B14P. The last codon of CYP2B14P exon 1 is a TAG translation stop codon, and thus CYP2B14P is a pseudogene. Three other inserts covering 34 kb of genomic sequence together carried 9 exons and the 5'- and 3'-flanking regions of the gene designated CYP2B16P. The 5'-splice site of CYP2B16P intron 1 is inactivated by the replacement of G by T in the normally invariant GT dinucleotide. Thus, CYP2B16P also has the characteristics of a pseudogene. The identification of CYP2B14P and CYP2B16P brings to at least seven the number of genes or pseudogenes shown by nucleotide sequence analysis to belong to the rat CYP2B subfamily. BIOCHEM PHARMACOL 52;6:963–965, 1996.

KEY WORDS. pseudogenes; phenobarbital; induction; CYP2B subfamily; cytochrome P450; constitutive

The CYP† superfamily of hemoproteins is involved in the oxidation of endogenous substrates such as steroids and fatty acids, as well as exogenous substrates such as drugs, mutagens, and carcinogens [1, 2]. A characteristic CYP feature is the presence of forms inducible by xenobiotics such as PB or methylcholanthrene, coexisting with constitutive forms [2]. Mammalian CYP proteins are classified according to their amino acid sequences into 14 families and 26 subfamilies [3].

The two major hepatic forms of the rat CYP2B subfamily, CYP2B1 and CYP2B2, are strongly induced by PB [4]. We recently isolated and characterized a third gene in the CYP2B subfamily, CYP2B3 [5], which like CYP2B1 and CYP2B2 [6, 7] had 9 exons and 8 introns. CYP2B3 is transcribed into a constitutive hepatic mRNA encoding a protein the deduced 491-amino acid sequence of which has 77% identity with the CYP2B1/CYP2B2 proteins [8, 9]. While cloning CYP2B3, we isolated several variant genomic inserts with restriction patterns different from those corresponding to CYP2B3, suggesting the existence of new genes [5]. We report here the further characterization of

MATERIALS AND METHODS Structure of CYP2B14P and CYP2B16P Genomic Inserts

To clone and characterize CYP2B3, a λEMBL4 rat genomic library was screened with probes from the 5', central, and 3' portions of the CYP2B3 cDNA [5]. Five variant inserts were obtained from the screen with the 5' CYP2B3 cDNA probe, all of which hybridized with CYP2B3 exon 1 probes, but not with probes for other CYP2B3 exons. Screening with the central CYP2B3 probe yielded B18, a variant insert that hybridized with probes for CYP2B3 exons 2 to 6. Finally, B2, a variant insert hybridizing with probes for CYP2B3 exons 7 to 9, was isolated by screening with the 3' CYP2B3 probe.

RESULTS AND DISCUSSION

Restriction mapping showed that the five 5' inserts overlapped each other. They all carried the 5'-flanking region, exon 1, and part of intron 1 of a CYP2B gene later designated CYP2B14P (Fig. 1A). B18 and B2 were also found to overlap. Together they carried exons 2 to 9 and the 3'-flanking region of a CYP2B gene later designated CYP2B16P (Fig. 1B). However, no overlap was found be-

these inserts, which led to the identification of two apparent pseudogenes, CYP2B14P and CYP2B16P.

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[†] Abbreviations: CYP, cytochrome P450; and PB, phenobarbital. Received 13 December 1995; accepted 25 April 1996.

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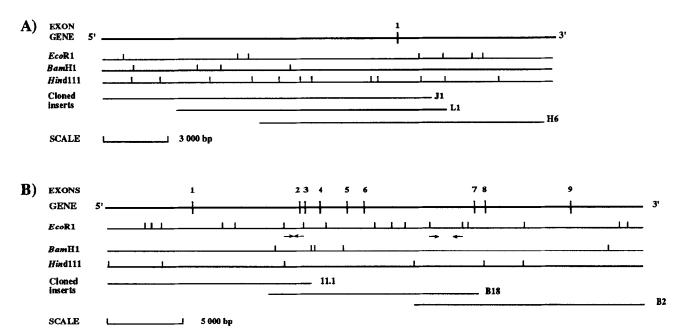


FIG. 1. Restriction maps of CYP2B14P (A) and CYP2B16P (B) including the lengths of representative cloned inserts. Methods of screening and analysis of genomic clones as well as restriction mapping and DNA sequencing have been described [5]. In addition to the 5', central, and 3' CYP2B3 cDNA probes described previously [5], a 450-bp AccI genomic fragment of insert B18 from CYP2B16P intron 1 was used to screen the genomic library to obtain clone 11-1. The overlaps in CYP2B16P introns 1 and 6 were confirmed by sequence analysis (see horizontal arrows in the figure). Nucleotide sequences were analysed using the University of Wisconsin GCG programs [10].

tween the five 5' inserts and B18. Be rescreening the genomic library with a CYP2B16P intron 1 probe, we isolated a clone the insert of which (11-1) overlapped with B18 but not with the 5' inserts. Furthermore, 11-1 carried exons 1, 2, and 3 as well as the 5'-flanking region of CYP2B16P (Fig. 1B), thus demonstrating that CYP2B14P and CYP2B16P are indeed two distinct genes. Insofar as comparisons are possible, the CYP2B14P and CYP2B16P restriction maps appear to be different from those reported for other cloned rat CYP2B genes [5-7, 11-13].

Sequence Analysis of CYP2B14P and CYP2B16P

By DNA sequence analysis, the last codon of CYP2B14P exon 1 was found to be a TAG stop codon, the presence of which was confirmed in three independently isolated cloned inserts. This indicates that CYP2B14P is a pseudogene. The nucleotide sequence of CYP2B16P exons 1 to 9, the intronic boundaries, and the 5'- and 3'-flanking regions were determined. The eight introns were found to be inserted at positions precisely equivalent to those of other CYP family 2 genes [5-7, 14, 15], except that the first codon of CYP2B16P exon 6 is deleted. The 5'-splice site of CYP2B16P intron 1 is abolished by the replacement of G by T in the normally invariant GT dinucleotide. Mutational replacement of this G by T or A completely inactivates the 5'-splice site of the human \(\beta\)-globin gene [16, 17], and no functional 5'-splice sites beginning with TT were found among more than 7500 natural splice sites examined [18,

19]. This indicates that CYP2B16P cannot direct the synthesis of a typical CYP2B protein and thus has the properties of a pseudogene, although the formal possibility that a nearby cryptic splice site may be functional cannot be excluded. The Genbank accession numbers for CYP2B14P and CYP2B16P are U33540, U33541, U33542, U33543, U33544, U33545, and U33546.

The nine CYP2B16P exons and the single cloned CYP2B14P exon define open reading frames encoding virtual proteins of 490 and 56 amino acids, respectively. The virtual CYP2B16P protein shares 81% identity with CYP2B1 and 75% with CYP2B3, while the virtual peptide encoded by CYP2B14P shares 70, 82, and 77% identity with the peptides encoded by exon 1 of CYP2B1, CYP2B3, and CYP2B16P, respectively. Thus, CYP2B14P and CYP2B16P are new members of the CYP2B subfamily.

Previous work led to the cloning and characterization at the DNA sequence level of all nine exons of four rat CYP2B genes [CYP2B1 [6], CYP2B2 [7, 12], CYP2B3 [5, 20], and CYP2B15 [21] (which appears to be the same as the gene designated CYP2B8 by Giachelli et al. [13])] as well as two gene fragments (CYP2B12 [12, 22] and gene 3 of Atchison and Adesnik [12, 20]). By sequence, CYP2B16P is different from all six of these, and CYP2B14P is different from at least five (since their cloned portions do not overlap, CYP2B14P and gene 3 may be the same gene). Thus, the present cloning of CYP2B14P and CYP2B16P brings to at least seven the number of sequenced rat CYP2B genes or gene fragments. Other rat CYP2B genes have been cloned [11, 13] but have yet to be characterized.

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References

- Guengerich FP (Ed.), Mammalian Cytochromes P-450, Vol. I. CRC Press, Boca Raton, FL, 1987.
- Gonzalez FJ, The molecular biology of cytochrome P450s. Pharmacol Rev 40: 243–288, 1988.
- Nelson DR, Koymans L, Kamataki T, Stegeman JJ, Feyereisen R, Waxman DJ, Waterman MR, Gotoh O, Coon MJ, Estabrook RW, Gunsalus IC and Nebert DW, P450 superfamily: Update on new sequences, gene mapping, accession numbers, and nomenclature. *Pharmacogenetics* 6: 1–42, 1996.
- Waxman DJ and Azaroff L, Phenobarbital induction of cytochrome P-450 gene expression. Biochem J 281: 577–592, 1997
- Jean A, Reiss A, Desrochers M, Dubois S, Trottier E, Trottier Y, Wirtanen L, Adesnik M, Waxman DJ and Anderson A, Rat liver cytochrome P450 2B3: Structure of the CYP2B3 gene and immunological identification of a constitutive P450 2B3-like protein in rat liver. DNA Cell Biol 13: 781–792, 1994.
- Suwa Y, Mizukami Y, Sogawa K and Fujii-Kuriyama Y, Gene structure of a major form of phenobarbital-inducible cytochrome P-450 in rat liver. J Biol Chem 260: 7980–7984, 1985.
- Mizukami Y, Sogawa K, Suwa Y, Muramatsu M and Fujii-Kuriyama Y, Gene structure of a phenobarbital-inducible cytochrome P-450 in rat liver. Proc Natl Acad Sci USA 80: 3958–3962, 1983.
- 8. Affolter M, Labbé D, Jean A, Raymond M, Noël D, Labelle Y, Parent-Vaugeois C, Lambert M, Bojanowski R and Anderson A, cDNA clones for liver cytochrome P-450s from individual Aroclor-treated rats: Constitutive expression of a new P-450 gene related to phenobarbital-inducible forms. DNA 5: 209–218, 1986.
- Labbé D, Jean A and Anderson A, A constitutive member of the rat cytochrome P450IIB subfamily: Full-length coding sequence of the P450IIB3 cDNA. DNA 7: 253–260, 1988.
- 10. Devereux J, Haeberli P and Smithies O, A comprehensive set

- of sequence analysis programs for the VAX. *Nucleic Acids Res* 12: 387–395, 1984.
- 11. Mizukami Y, Fujii-Kuriyama Y and Muramatsu M, Multiplicity of deoxyribonucleic acid sequences with homology to a cloned complementary deoxyribonucleic acid coding for rat phenobarbital-inducible cytochrome P-450. *Biochemistry* 22: 1223–1229, 1983.
- 12. Atchison M and Adesnik M, A cytochrome P-450 multigene family. Characterization of a gene activated by phenobarbital administration. *J Biol Chem* **258**: 11285–11295, 1983.
- Giachelli CM, Lin-Jones J and Omiecinski CJ, Isolation and characterization of rat cytochrome P-450IIB gene family members. J Biol Chem 264: 7046–7053, 1989.
- Morishima N, Yoshioka H, Higashi Y, Sogawa K and Fujii-Kuriyama Y, Gene structure of cytochrome P-450(M-1) specifically expressed in male rat liver. *Biochemistry* 26: 8279– 8285, 1987.
- Chan G and Kemper B, Structure of the rabbit cytochrome P450IIC3 gene, a constitutive member of the P450IIC subfamily. Biochemistry 29: 3743–3750, 1990.
- Treisman R, Orkin SH and Maniatis T, Specific transcription and RNA splicing defects in five cloned β-thalassemia genes. Nature 302: 591–596, 1983.
- 17. Kazazian HH Jr, Orkin SH, Antonarakis SE, Sexton JP, Boehm CD, Goff SC and Waber PG, Molecular characterization of seven β-thalassemia mutations in Asian Indians. EMBO J 3: 593–596, 1984.
- 18. Senapathy P, Shapiro MB and Harris NL, Splice junctions, branch point sites, and exons: Sequence statistics, identification, and application to the genome project. *Methods Enzymol* **183:** 252–278, 1990.
- 19. Jackson IJ, A reappraisal of non-consensus mRNA splice sites. *Nucleic Acids Res* 19: 3795–3798, 1991.
- Atchison M and Adesnik M, Gene conversion in a cytochrome P-450 gene family. Proc Natl Acad Sci USA 83: 2300– 2304, 1986.
- Nakayama K, Suwa Y, Mizukami Y, Sogawa K and Fujii-Kuriyama Y, Cloning and sequencing of a novel rat cytochrome P450 2B-encoding gene. Gene 136: 333–336, 1993.
- 22. Friedberg T, Grassow MA, Bartlomowicz-Oesch B, Siegert P, Arand M, Adesnik M and Oesch F, Sequence of a novel cytochrome CYP2B cDNA coding for a protein which is expressed in a sebaceous gland, but not in the liver. *Biochem J* 287: 775–783, 1992.